BAX® System Real-Time PCR Assay  
E. coli O157:H7

Part D14203648

INTENDED USE

Food processors and associated laboratories can use the BAX® System as a quick and reliable method for detecting E. coli O157:H7 in ground beef, beef trim, spinach and lettuce. The real-time PCR assay was designed to report yes/no results for E. coli O157:H7, a recognized foodborne pathogen with a low infectious dose. With a processing time of approximately 55 minutes in the BAX® System Q7 instrument, results are available in less than 11 hours for ground beef, less than 12 hours for beef trim and less than 10 hours for spinach and lettuce. BAX® Systems are designed for use by qualified personnel who follow standard microbiology laboratory practice, including the safe handling and disposal of potentially pathogenic materials.

PRINCIPLE OF THE METHOD

The BAX® System uses the Polymerase Chain Reaction (PCR) to amplify specific DNA fragments, which are stable and unaffected by growth conditions. Each fragment is a genetic sequence that is unique to the targeted organism, thus providing a highly reliable indicator that the organism is present. The BAX® System simplifies the PCR process by combining the requisite PCR reagents (primers, polymerase, nucleotides and internal positive controls) into a stable, dry, manufactured tablet already packaged inside the PCR tubes. After hydrating these tablets with prepared samples, the tubes remain sealed to reduce the potential for contamination.

In a typical PCR application, sample DNA is combined with DNA polymerase, nucleotides and primers that are specific for a given nucleotide sequence. The mixture then undergoes a series of timed heating and cooling cycles. Heating denatures the DNA, separating it into single strands. As the mixture cools, the primers recognize and anneal (bind) to the targeted DNA sequence. DNA polymerase then uses nucleotides to extend the primers, thus creating two copies of the targeted fragment (amplification). Repeating cycles of denaturing, annealing and extending produces an exponential increase in the number of target DNA fragments, creating millions of copies in a very short time. If the target sequence is not present, no detectable amplification takes place.

The BAX® System PCR tablets used in this assay contain multiple target-specific, dye labeled probes. Probes are short oligonucleotides with a quencher dye at one end that greatly reduces fluorescence from the fluorophore dye at the opposite end. During PCR, probes bind to a specific area within the targeted sequence. The fluorophore is then quenched, allowing for increased fluorescent signal. The BAX® System Q7 instrument uses dye-specific filters to measure signal at the end of each cycle and report positive/negative results for each target.

STORAGE AND SHELF LIFE

Reagents and PCR tubes with tablets should be kept refrigerated at 2–8°C. Do not freeze. Reagents should be used by the expiration date stamped on the individual labels. After protease has been added to the lysis buffer, shelf life of the solution is 2 weeks when stored at 2-8°C.

INSTRUMENT REQUIREMENTS

This assay can be used only on a BAX® System Q7 instrument running software v2.7 or higher. After installing the software but before running the real-time E. coli O157:H7 assay for the first time, run a calibration report to check that “Real Time E. coli O157:H7” appears in the list of calibration files. If it does not, you will need to re-calibrate the Q7 instrument to load the required dyes. Be sure to allow enough time to complete the calibration (about 1.5 to 2 hours) before starting the assay. For instructions and tips on calibrating the instrument, see the BAX® System User Guide.

STANDARD PROTOCOL FOR ENRICHED SAMPLES

1. Prepare enrichment broth

1.1 Dissolve 22.5 g BAX® system enrichment media for E. coli O157:H7 MP in 1 L distilled water and mix. Do not boil. Adjust pH to a final value of 7.2±0.2, then autoclave at 121°C for 15 minutes.

Note: DuPont™ StatMedia™ soluble packets may also be used to prepare E. coli O157:H7 enrichment broth. See instructions on packet or in User Guide.

2. Collect and enrich samples

2.1 Prepare samples for enrichment.

- Ground beef: Stomach 65 g sample with 585 mL pre-warmed (42°C) BAX® system enrichment broth.
- Beef trim: Mix 375 g sample by hand with 1.5 L pre-warmed (45°C) BAX® system enrichment broth.
- Spinach and lettuce: Stomach 25 g sample with 225 mL pre-warmed (42°C) BAX® system enrichment broth.

Stomacher with bags

Incubator capable of maintaining 42±2°C

*Cluster tubes with caps and racks

*Tips for all pipettes

*Powder-free nitrile gloves

*Sufficient supply for 96 tests included in BAX® System start-up package

3. Prepare equipment

3.1 Turn on the heating blocks to 37±2°C and 95±3°C.

3.2 Make sure cooling blocks are chilled to 2-8ºC.

3.3 Power on the Q7 instrument and launch the BAX® system application.

3.4 Create a rack file (see User Guide for details).

3.5 Initialize the instrument by selecting RUN FULL PROCESS from the OPERATION menu.

4. Perform lysis

4.1 Label and arrange cluster tubes in rack according to the rack file.

4.2 Mix lysis reagent by adding 150 µL protease to a 12-ml bottle of lysis buffer.

4.3 Transfer 200 µL lysis reagent to each cluster tube.

4.4 Transfer 20 µL enriched sample to each cluster tube.

4.5 Heat at 37±2°C for 20 minutes.

4.6 Heat at 95±3°C for 10 minutes.

4.7 Cool for 5 minutes in cooling block.

5. Hydrate PCR tablets

5.1 Place a chilled PCR tube insert into cooling block and cover with PCR tube holder.

5.2 Arrange strips of PCR tubes according to your rack file.

5.3 Remove the caps from the first strip of tubes with the decapping tool.

5.4 Transfer 30 µL lystate into PCR tubes, then seal with flat optical caps.

Note: PCR tablets must be hydrated and re-sealed within 10 minutes after removing the caps from the PCR tubes.

6. Amplify and detect

6.1 At the “Ready for Rack Load” prompt, click the NEXT button and open the instrument drawer.

6.2 Place the rack of PCR tubes over the wells in the drawer, and check that the tubes are seated correctly.

6.3 Close the drawer, and click the NEXT button to begin automated processing.

Field of use: Data obtained from the BAX® System should not be used for human diagnostic or treatment purposes. Equipment is not approved by the United States Food and Drug Administration or any other U.S. or non-U.S. regulatory agency for use in human diagnostics or treatment. The BAX® System should not be used as the sole basis for assessing the safety of products for release to consumers. The information generated is only to be used in conjunction with the user’s regular quality assurance program. Not approved for clinical diagnosis. Use for research and development, quality assurance and quality control under supervision of technically qualified persons.

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7. Review results

Qualicon is confident that the BAX® System possesses all of the characteristics described on its wall of accolades:

- For all assays present in the sample, the well is red with a “plus” sign (positive).
- For any other results present in the sample, the well is green with a “minus” sign (negative).

The BAX® System Real-Time PCR Assay for E. coli O157:H7 has been internally validated on samples of ground beef, beef trim, spinach, and green lettuce. Additional data will be submitted to the AOAC Research Institute for Performance Tested Method certification.

CONFIRMATION

If desired, ground beef and trim samples can be confirmed according to the USDA-FSIS standards by performing an IMS test and stepping presumptive E. coli O157:H7 cells onto Rainbow® Agar O157 (Biolog Inc., Hayward, California). In addition, directly streak 1 ul. enrichment onto Rainbow® Agar O157 without IMS to confirm rough E. coli O157:H7 strains which may not be captured by IMS. Incubate all plates for 18-24 hours at 37°C. Spinach and lettuce samples can be confirmed according to FDA standards by streaking 10 ul. enrichment onto TCSCM plates and incubating for 18-24 hours at 35-37°C. In addition, for samples with high levels of background flora, perfom an IMS step before streaking presumptive E. coli O157:H7 cells onto TCSCM plates. For all confirmation procedures, typical isolates should be confirmed with appropriate biochemical and serological methods.

DISPOSAL

Decontaminate materials and dispose of biohazardous waste according to your site practices and as required by federal, state and local regulations. If you have questions about proper waste disposal at your site for the materials provided by DuPont Qualicon, please call 1-800-863-6842, fax 302-351-6454, or visit www.qualicon.com.

LIMITATION OF WARRANTY AND LIABILITY

NOTICE: READ THIS LIMITATION AND LIABILITY BEFORE USING THE BAX® SYSTEM EQUIPMENT, ASSAYS, AND/OR MEDIA (“BAX® SYSTEM”). If the terms are not acceptable, notify DuPont Qualicon (“Qualicon”) immediately and arrangements will be made for return of the unused equipment, assays, and/or media to Qualicon and for the refund of the purchase price, less shipping cost.

1. BAX® system equipment should only be used with BAX® system assays.
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5. Externally caused abuse, misuse, negligence or improper operation are not covered by this warranty. Equipment failures caused by spills, abuse, misuse, negligence, or improper operation are not covered by this warranty. Modifications, service or repairs by parties other than Qualicon-authorized providers are not covered by this warranty and, in fact, void this warranty. Circumstances beyond the reasonable control of Qualicon, including fire, explosions, accidents, flood, labor trouble or shortage, war, act of or authorized by any government, inability to obtain suitable material, equipment, fuel, power or transportation, or acts of God are not covered under this warranty.

6. The BAX® system is designed to test only for the presence of the target organisms specified in the particular assay. The BAX® system has been tested against many, but not all, strains of the target within the sample types specified in the user documentation. Qualicon, therefore, cannot and does not make any representation or warranty that the BAX® system is capable of detecting every organism in the target genus, serotype, or species in any sample source. Accordingly, the BAX® system should not be used as the sole test for the release of user’s products, nor should it be used as the sole basic test for determining the safety of user’s products.

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