Vibrio are frequently found in coastal waters and other brackish environments, and pathogenic species can cause gastrointestinal illness when ingested by humans. Foodborne infection is typically associated with eating raw or undercooked fish and shellfish, especially oysters. For seafood processors and inspectors, current culture methods require at least 3-5 days for results and subjective interpretation. The BAX® system Vibrio assay, however, delivers differentiated results that are reliable and highly specific in less than 24 hours.

### Benefits of the BAX® System Real-Time PCR Assay
- **Speed** - Reliable results within 24 hours
- **Accuracy** - clear and reproducible results, independent of operator technique
- **Ease of use** - minimal hands-on time, tableted PCR reagents and automated processing and analysis reduce operator error
- **Convenience** - pre-packaged PCR reagent tablets provide consistency, stability and long shelf-life
- **Support** – customer-focused dependability from DuPont Qualicon to answer your questions and keep your operation running smoothly

### Features
- Up to 96 enriched samples processed in about one hour
- Clear, reproducible yes-or-no results for *V. cholerae*, *V. parahaemolyticus* and *V. vulnificus*
- Reliably detects $10^4$ cfu/mL
- Excellent inclusivity/exclusivity for all three target species
- Internally validated on oysters, shrimp, scallops, crab and tuna
- LIMS-compatible electronic data for easy storage, sharing and retrieval
- Can be run with other BAX® system real-time assays
- Includes all lysis reagents needed for processing
- Step-by-step instructions in each package
- Will be submitted to AOAC-RI for Performance Tested Methods™ status
**Sample preparation**

Stomach or blend 25 g sample with 225 mL alkaline peptone water.

When using Oxoid CM1117 use at half strength (20g/Litre).

Incubate at 35°C for 18-20 hours.

**BAX® system protocol**

8:00 Create rack file and warm up cycler.

8:05 Mix protease with lysis buffer and transfer 200 µL of lysis reagent to cluster tubes.

8:10 Transfer 5-µL samples to cluster tubes.

8:20 Heat cluster tubes for 20 minutes at 37°C, then 10 minutes at 95°C.

8:50 Cool cluster tubes for 5 minutes in cooling block, then transfer 30 uL to PCR tubes in cooling block.

9:00 Place sealed PCR tubes in cycler and run program.

10:05 Review results.

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